located at the C-terminus of the Osterix protein. The antibodies were affinity purified over a 3M Emphaze Biosupport Medium AB1 column (Pierce) coupled to the 14-amino acid peptide and were eluted at low and high pH. They were then dialysed against Tris-buffered saline. --

The present amendments correct inadvertent typographical errors. No new matter is introduced by these amendments.

## II. RESPONSE TO NOTICE

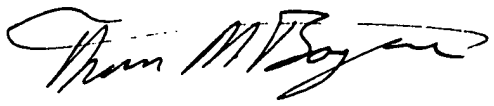
The Notice indicates that the specification contains reference to a SEQ ID NO: 9 on page 30, yet only SEQ ID NO: 1 through SEQ ID NO: 6 are provided. The Notice therefore concludes that SEQ ID NO: 7 and 8 are missing from the sequence listing as filed. Applicants have identified the reference to SEQ ID NO: 9 as a typographical error. Applicants have amended the passage of the specification to properly refer to SEQ ID NO: 6. Therefore, no SEQ ID NO: 7 nor SEQ ID NO: 8 are required.

The Notice indicates that the specification lists a 14 amino acid peptide on page 103 without the identification of a corresponding sequence listing. Applicants have amended the specification to insert the proper reference to SEQ ID NO: 3, which was inadvertently omitted in the specification as filed. All listed sequences in the specification are now believed to properly refer to the appropriate sequence listings.

The Notice provides no other reasons for non-compliance with the requirements of sequence listings. A copy of the Notice is enclosed. Applicants respectfully submit that the above amendments are fully responsive to the Notice in that the sequence listing itself does not appear to contain errors or other, non-complying content.

Please date stamp and return the accompanying postcard to evidence receipt of these documents.

Respectfully submitted,



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Date: July 17, 2002

## Appendix A:

**Marked up version of replacement paragraphs of amendments t the specification showing all changes made. Insertions are underlined and deletions are within square brackets.**

On page 29:

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template or where one seeks to isolate Osterix encoding sequences from related species, functional equivalents, or the like, less stringent hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ conditions such as 0.15M-1.0M salt, at temperatures ranging from 20°C to 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In fact, the inventors have been able to detect a human equivalent for mouse Osterix by Southern hybridization of human cDNA with a sequence of mouse Osterix (SEQ ID NO:[9]6) under a low stringency condition (1M NaCl, 30-45% formamide, 10% dextran sulfate, at 37°C). In any case, it is generally appreciated that conditions can be rendered more stringent by decreasing NaCl concentrations or by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

On page 103:

**Purification of anti Osterix antibodies.** Antibodies were created by immunizing rabbits with a 14-amino acid peptide (AHGGSPEQSNLLEI; SEQ ID NO: 3) located at the C-terminus of the Osterix protein. The antibodies were affinity purified over a 3M Emphaze Biosupport Medium AB1 column (Pierce) coupled to the 14-amino acid peptide and were eluted at low and high pH. They were then dialysed against Tris-buffered saline.